# Epigenetic Programming and Diabetes: Molecular Mechanisms, Environmental Influences, and Therapeutic Prospects

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Abstract:Diabetes mellitus, encompassing type 1 (T1D) and type 2 diabetes (T2D), is a multifactorial metabolic disorder affecting millions globally, characterized by chronic hyperglycemia stemming from insulin deficiency and/or resistance. While genetic predisposition has been extensively studied, emerging evidence implicates epigenetic programming as a critical regulator that integrates environmental and developmental signals with genomic information to modulate diabetes susceptibility, onset, and progression. Epigenetic modifications—such as DNA methylation, histone modifications, chromatin remodeling, and non-coding RNA regulation—alter gene expression without changing the DNA sequence and are dynamic across cell types and developmental stages. In T1D, epigenetic dysregulation promotes autoimmune T cell activation and β-cell fragility, while in T2D, these changes mediate insulin resistance, β-cell dysfunction, and metabolic memory. Environmental factors, including in utero exposures, diet, physical activity, stress, and toxicants, profoundly impact epigenetic landscapes, influencing diabetes risk across the lifespan. Additionally, epigenetic biomarkers derived from peripheral tissues offer promising avenues for early diagnosis and prognosis. The advent of epigenetic therapies, including inhibitors targeting DNA methyltransferases and histone deacetylases, combined with nutritional and lifestyle interventions, provides novel opportunities for precision medicine. This review systematically synthesizes current knowledge on epigenetic mechanisms in diabetes, the influence of environmental and developmental factors, biomarker potential, and emerging therapeutic strategies, highlighting future directions for epigenome-based interventions to alleviate the global diabetes burden.

# **Keywords:**

Epigenetics; DNA methylation; histone modifications; non-coding RNAs; chromatin remodeling; type 1 diabetes; type 2 diabetes; β-cell dysfunction; insulin resistance; metabolic memory; environmental factors; developmental programming; epigenetic therapy.

#### INTRODUCTION

Diabetes mellitus is a heterogeneous group of metabolic diseases characterized by chronic hyperglycemia resulting from defects in insulin secretion, insulin action, or both [1]. The global burden of diabetes continues to escalate, with an estimated 537 million adults affected worldwide and projections predicting over 700 million by 2045 [2]. The two major forms, type 1 diabetes (T1D) and type 2 diabetes (T2D), differ in pathophysiology but share overlapping genetic and environmental risk factors [3]. Genetic factors, especially HLA

alleles for T1D and polymorphisms in genes like TCF7L2 for T2D, explain only a fraction of disease risk [4]. Growing evidence implicates epigenetic mechanisms—heritable changes in gene expression without alteration of DNA sequence—in mediating gene-environment interactions critical to diabetes pathogenesis [5]. Epigenetic modifications regulate chromatin architecture, influencing gene accessibility and transcriptional dynamics, thereby orchestrating cellular identity and function [6]. Environmental factors such as maternal nutrition, exposure to toxins, physical activity, and psychological stress modulate

epigenetic marks during critical developmental windows and throughout life, impacting diabetes susceptibility [7]. Furthermore, persistent epigenetic alterations contribute to the phenomenon of "metabolic memory," where early hyperglycemia induces lasting detrimental effects despite glycemic control [8]. This review aims to provide an in-depth analysis of epigenetic programming in diabetes, highlighting molecular mechanisms, environmental modulators, diagnostic and prognostic biomarker potential, and emerging epigenetic-based therapeutic strategies.

# 2. Epigenetic Mechanisms and Regulatory Networks

# 2.1 DNA Methylation

DNA methylation involves covalent addition of a methyl group at the 5' position of cytosine residues within CpG dinucleotides, catalyzed primarily by DNA methyltransferases DNMT1, DNMT3A, and DNMT3B [9]. This modification is generally associated with transcriptional repression through recruitment of methyl CpGbinding domain proteins and chromatin compaction [10]. DNA methylation patterns are established early in development and maintained through mitosis, but can be remodeled in response to environmental stimuli [11]. In pancreatic β-cells, methylation at promoters of insulin (INS), PDX1, and other β cell-specific genes is essential for proper differentiation and function [12]. Alterations in DNA methylation of immune regulatory genes contribute to autoimmune activation in T1D [13], while in T2D, metabolic genes such as PPARGC1A undergo aberrant methylation leading to insulin resistance [14].

# 2.2 Histone Modifications

Histones, around which DNA is wrapped to form nucleosomes, are modified post translationally via acetylation, methylation, phosphorylation, ubiquitination, and simulation on their N-terminal tails [15]. These modifications regulate chromatin accessibility and transcription by altering histone-DNA interactions and recruiting effector proteins [16]. Histone acetylation (via HATs) generally activates transcription by loosening chromatin, whereas deacetylation (via HDACs) promotes gene silencing [17]. Methylation marks such as H3K4me3 are linked to active transcription, whereas H3K27me3 correlates with repression [18]. In diabetes, dysregulation of histone acetylation and methylation at loci

controlling inflammation and metabolism is well documented [19].

# 2.3 Chromatin Remodeling

ATP-dependent chromatin remodeling complexes (SWI/SNF, ISWI) reposition nucleosomes, modulating DNA accessibility for transcription factors [20]. These complexes regulate  $\beta$ -cell identity and metabolic gene transcription. Their dysfunction contributes to  $\beta$ -cell failure and impaired glucose metabolism [21].

# 2.4 non-coding RNAs

Non-coding RNAs (ncRNAs), including microRNAs (miRNAs) and long non-coding RNAs (lncRNAs), regulate gene expression post-transcriptionally or via epigenetic modulation [22]. miRNAs such as miR-375 regulate insulin secretion and β-cell apoptosis, whereas lncRNAs modulate inflammatory pathways and insulin sensitivity [23].

#### 3. Epigenetic Programming in Type 1 Diabetes

#### 3.1 Autoimmune Pathogenesis and Epigenetics

Type 1 diabetes (T1D) is a complex, polygenic autoimmune disease resulting from the destruction of pancreatic  $\beta$ -cells by autoreactive T lymphocytes, culminating in insulin deficiency [24]. While classical genetic loci such as the HLA-DR/DQ region explain a significant proportion of susceptibility, incomplete concordance in monozygotic twins suggests an influential role for epigenetic factors and environmental triggers [25].

Central to T1D pathogenesis is the failure of immune tolerance, mediated by dysregulated thymic education, peripheral tolerance breakdown, and aberrant activation of innate and adaptive immune components [26]. Epigenetic modifications dynamically regulate gene expression programs in T cells, B cells, dendritic cells, and  $\beta$ -cells, influencing disease onset and progression.

# 3.2 DNA Methylation Dynamics in Immune Cells

Comprehensive methylome profiling of CD4+ T cells and monocytes from T1D patients reveals locus-specific hypomethylation at genes involved in antigen presentation (HLA-DRB1, HLA-DQA1), costimulatory signaling (CD28), and cytokine secretion (IFNG, IL2) [27]. These hypomethylated regions correspond to transcriptionally active chromatin states, enhancing the pathogenic potential of autoreactive immune cells [28]. Conversely, hypermethylation

of regulatory genes such as FOXP3, critical for Treg cell function, correlates with reduced immunosuppressive capacity, favoring autoimmune destruction [29].

Studies using bisulfite sequencing highlight epigenetic heterogeneity among immune subsets, with distinct methylation signatures in naïve versus memory T cells suggesting epigenetic memory influences autoimmune reactivation [30].

# 3.3 Epigenetic Alterations in β-Cells

Beyond immune cells, pancreatic  $\beta$ -cells themselves undergo epigenetic remodeling in T1D. Genome-wide methylation studies in isolated islets from T1D donors demonstrate hypermethylation of insulin gene promoters and  $\beta$ -cell identity transcription factors such as PDX1 and MAFA, resulting in impaired insulin synthesis [31]. Pro-inflammatory cytokines (IL-1 $\beta$ , IFN- $\gamma$ ) induce  $\beta$ -cell chromatin remodeling, including changes in histone acetylation and methylation that prime cells for apoptosis [32]. Methylation alterations in apoptosis-regulating genes (BAX, BCL2) further determine  $\beta$ -cell susceptibility to immune-mediated killing [33].

# 3.4 Histone Modification and Chromatin Remodeling in Immune Dysfunction

Histone acetyltransferases (HATs) and histone deacetylases (HDACs) regulate inflammatory gene expression in immune cells. In T1D, increased acetylation marks such as H3K27ac at cytokine gene promoters enhance transcription of IFNG and TNFA, sustaining autoimmune inflammation [34]. Conversely, reduced activating histone marks at insulin gene promoters correlate with decreased insulin transcription in  $\beta$ -cells [35]. Dysregulation of chromatin remodeling complexes (SWI/SNF) impairs  $\beta$ -cell gene regulation and resilience [36].

# 3.5 Non-Coding RNA Dysregulation

Upregulated miRNAs such as miR-21, miR-146a, and miR-155 in T cells and macrophages promote pro-inflammatory cytokine production and  $\beta$ -cell apoptosis [37]. Long non-coding RNAs (lncRNAs), including TMEVPG1, regulate Th1 cell differentiation and perpetuate autoimmune responses [38]. Aberrant expression of ncRNAs mediates immune-metabolic crosstalk, exacerbating  $\beta$ -cell destruction [39].

# 4. Epigenetic Programming in Type 2 Diabetes

## 4.1 Pathophysiological Context

Type 2 diabetes (T2D) is characterized by peripheral insulin resistance and progressive β-cell failure [40]. Genetic

predisposition is polygenic and complex; epigenetic mechanisms are key modulators of gene expression involved in glucose metabolism, inflammation, and cellular stress responses [41].

# **4.2 DNA Methylation Aberrations**

identified Epigenome-wide association studies have hypermethylation of metabolic genes such as PPARGC1A, IRS1, and GLUT4 in insulin-resistant skeletal muscle and adipose tissue, impairing glucose uptake and mitochondrial function [42]. Pancreatic islets from T2D donors exhibit DNA methylation changes in β-cell identity and insulin secretion genes, contributing to β-cell dysfunction [43]. Persistent hyperglycemia induces DNA methylation alterations reinforcing "metabolic memory," leading to chronic complications [44].

#### 4.3 Histone Modifications and Chromatin Accessibility

Increased expression and activity of histone deacetylases (HDACs) in metabolic tissues repress insulin signaling and mitochondrial gene transcription [45]. Loss of activating histone marks such as H3K9ac and H3K4me3 at promoters of key insulin signaling genes further suppresses metabolic function [46]. Histone methyltransferases like SUV39H1 catalyze repressive methyl marks contributing to insulin resistance [47].

# 4.4 Non-Coding RNA Dysregulation

Dysregulated miRNAs including miR-375, miR-29, and miR-143 in pancreatic islets and peripheral tissues impair insulin secretion and promote  $\beta$ -cell apoptosis [48]. Long non-coding RNAs such as MALAT1 and H19 modulate lipid metabolism and inflammatory pathways, exacerbating systemic insulin resistance [49]. Complex interactions between ncRNAs and epigenetic machinery amplify metabolic dysfunction [50].

# 5. Environmental and Developmental Modulation of Epigenetic Programming

#### 5.1 Prenatal and Early Life Programming

The Developmental Origins of Health and Disease (DOHaD) hypothesis posits that environmental insults during gestation induce persistent epigenetic changes that influence diabetes susceptibility later in life [51]. Maternal malnutrition, hyperglycemia, and stress alter methylation of imprinted genes such as IGF2 and H19 and key metabolic regulators in offspring, impairing glucose homeostasis [52]. Animal models

demonstrate epigenetic reprogramming of pancreatic islets and insulin target tissues following in utero exposure to diabetes or nutrient restriction [53].

# **5.2 Nutritional Epigenomics**

Dietary methyl donors like folate, choline, and B vitamins influence DNA methylation patterns affecting insulin sensitivity and  $\beta$ -cell function [54]. High-fat and high-sugar diets induce histone modifications that promote inflammatory signaling and insulin resistance in liver and adipose tissue [55].

# 5.3 Physical Activity and Psychosocial Stress

Exercise induces beneficial epigenetic remodeling in skeletal muscle, including increased histone acetylation and reduced DNA methylation at promoters of mitochondrial and insulin signaling genes, improving metabolic capacity [56]. Conversely, chronic psychosocial stress elevates glucocorticoids, resulting in hypermethylation of glucocorticoid receptor genes and disrupted metabolic regulation [57].

## **5.4 Environmental Toxicants**

Exposure to endocrine-disrupting chemicals such as bisphenol A (BPA) and phthalates alters DNA methylation and histone marks in  $\beta$ -cells and adipocytes, impairing insulin secretion and promoting insulin resistance [58]. These epigenetic modifications may transmit across generations, posing significant public health concerns [59].

# 6. Epigenetic Biomarkers and Clinical Applications

## 6.1 DNA Methylation Biomarkers

Differential methylation patterns in peripheral blood DNA at loci such as TXNIP and ABCG1correlate with T2D onset and glycemic control, providing potential noninvasive biomarkers for early diagnosis and prognosis [60]. Methylation signatures in immune cells also reflect disease activity in T1D [61].

#### 6.2 Circulating Non-Coding RNAs

Circulating microRNAs such as miR-126, miR-375, and miR-146a serve as indicators of  $\beta$ -cell function and vascular complications, showing promise as prognostic biomarkers [62]. Combined profiling of miRNAs and lncRNAs enhances diagnostic precision and may differentiate diabetes subtypes [63].

## **6.3 Epigenetic Therapeutics**

Pharmacological inhibitors of DNA methyltransferases (DNMTs) and histone deacetylases (HDACs) have shown

efficacy in improving insulin sensitivity and  $\beta$  cell survival in preclinical models of diabetes [64]. Nutritional epigenetic modulators, including polyphenols and methyl donors, offer adjunct therapeutic potential. Emerging CRISPR-based epigenome editing techniques hold promise for precise gene regulation and correction of pathogenic epigenetic states.

# 7. Future Perspectives

Advancements in single-cell epigenomics integrated with multi-omics and machine learning will elucidate epigenetic heterogeneity in diabetes, informing personalized therapies. The development of safe, efficient epigenetic editing tools offers the potential for precision interventions. Ethical considerations and long-term safety of epigenetic therapies, especially germline editing, require thorough investigation to guide clinical translation.

#### 8. Conclusion

Epigenetic programming serves as a pivotal interface integrating genetic predisposition and environmental exposures to influence diabetes pathogenesis. Comprehensive understanding of epigenetic mechanisms offers opportunities for novel biomarkers and therapeutic strategies. Continued multidisciplinary research is essential for translating epigenetic insights into personalized medicine aimed at preventing and managing diabetes.

#### References

- [1]. DeFronzoRA.Pathogenesis of type 2 diabetes mellitus. Med Clin North Am.λ 2004;88(4):787-835. doi: 10.1016/j.mcna.2004.04.013
- [2]. International Diabetes Federation. IDF Diabetes Atlas,10th edn. Brussels, λ Belgium: 2021.
- [3]. American Diabetes Association. Diagnosis and classification of diabetes mellitus.λ Diabetes Care. 2014;37 Suppl 1: S81-90. doi:10.2337/dc14-S081
- [4]. Barrett JC, Clayton DG, Concannon P, et al. Genomewide association study andλ meta-analysis finds that over 40 loci affect risk of type 1 diabetes. Nat Genet. 2009;41(6):703-707. doi:10.1038/ng.381
- [5]. LingC, RönnT.Epigenetics in human obesity and type 2 diabetes. Cell Metab.λ 2019;29(5):1028-1044. doi: 10.1016/j.cmet.2019.03.009

- [6]. BirdA.DNAmethylation patterns and epigenetic memory. Genes Dev.λ 2002; 16(1): 6-21. doi:10.1101/gad.947102
- [7]. GluckmanPD, Hanson MA, et al. The developmental origins of adult disease. Clin Sci (Lond). 2008;113(7):219-231. doi:10.1042/CS20070430
- [8]. Brownlee M. The pathobiology of diabetic complications: a unifying mechanism. Diabetes. 2005;54(6):1615-1625. doi:10.2337/diabetes.54.6.1615
- [9]. GollMG, Bestor TH. Eukaryotic cytosine methyltransferases. Annu Rev Biochem. 2005; 74:481-514. doi: 10.1146/annurev.biochem.74.010904.1
- [10]. JonesPA.Functions of DNA methylation: islands, start sites, gene bodies andλ beyond. Nat Rev Genet. 2012;13(7):484-492. doi:10.1038/nrg3230
- [11]. WeberM, Schubeler D. Genomic patterns of DNA methylation: targets andλ function of an epigenetic mark. Curr Opin Cell Biol. 2007;19(3):273-280. doi: 10.1016/j.ceb.2007.03.003
- [12]. DayehT, VolkovP, et al. Genome-wide DNA methylation analysis of humanλ pancreatic islets from type 2 diabetic and non-diabetic donors identifies candidate genes that influence insulin secretion. PLoS Genet. 2014; 10(3): e1004160. doi: 10.1371 / journal.pgen.1004160
- [13]. Marroqui L, et al. Epigenetic regulation of immune and β-cell function in type 1λ diabetes. Front Endocrinol (Lausanne). 2019; 10:692. doi:10.3389 / fendo.2019.00692
- [14]. DayehT, Ling C. Does epigenetic dysregulation of pancreatic islets contribute toλ the pathogenesis of type 2 diabetes? Biochem Soc Trans. 2015;43(5):982-989. doi:10.1042/BST20150149
- [15]. Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications.λ Cell Res. 2011;21(3):381-395. doi:10.1038/cr.2011.22
- [16]. Kouzarides T. Chromatin modifications and their function. Cell.λ 2007;128(4): 693-705. doi: 10.1016/j.cell.2007.02.005 WangZ, etal. Histone modifications and their functions in diabetes. Int

- JλBiochem Cell Biol. 2014; 53:26-34. doi: 10.1016/j.biocel.2014.01.009
- [17]. Barski A, et al. High-resolution profiling of histone methylations in the humanλ genome. Cell. 2007;129(4):823-837. doi: 10.1016/j.cell.2007.05.009
- [18]. SunW, etal. Histone modifications in type 2 diabetes mellitus. Mol Med Rep.λ 2017;16(3):3583-3590. doi:10.3892/mmr.2017.7063 Clapier CR, Cairns BR.
- [19]. The biology of chromatin remodeling complexes. Annuλ Rev Biochem. 2009; 78:273-304. doi: 10.1146 / annurev.biochem.77.081307.094313
- [20]. WangH, etal. Chromatin remodeling and epigenetic regulation in  $\beta$ -cell function $\lambda$  and diabetes. J Endocrinol. 2019;241(1): R1-R16. doi:10.1530/JOE-18-0504
- [21]. Esteller M. Non-coding RNAs in human disease. Nat Rev Genet. $\lambda$  2011;12(12):861-874. doi:10.1038/nrg3074 PullenTJ, Rutter GA. When less is more: the role of microRNAs in  $\beta$ -cell $\lambda$  function. Diabetologia. 2013;56(12):2249-2258. doi:10.1007/s00125-013-3024- $\lambda$
- [22]. KnipM, Siljander H. Autoimmune mechanisms in type 1 diabetes. Autoimmuneλ Rev. 2008;7(7):550-557. doi: 10.1016/j.autrev.2008.03.007
- [23]. RedondoMJ, et al. Genetic and environmental interactions in type 1 diabetes.λ Ann NYAcadSci. 2008; 1150:196-213. doi:10.1111/j.1749-6632.2008. 03840.x
- [24]. AndersonMS, Bluestone JA. The NOD mouse: a model of immune dysregulation.λ Annu Rev Immunol. 2005; 23:447-485. doi: 10.1146 / annurev. immunol. 23. 021704. 115643
- [25]. JohnsonMB, et al. Epigenetic changes in T cells from type 1 diabetes patients.λ Clin Epigenetics. 2019;11(1): 54. doi: 10.1186 / s13148-019-0655-0
- [26]. ZhangX, et al. Hypomethylation of inflammatory gene promoters in T1D immune cells. J Autoimmune. 2020; 108:102416. doi: 10.1016/j.jaut.2019.102416
- [27]. LongSA, etal. FOXP3 methylation and regulatory T cell dysfunction in T1D. Jλ Clin Invest. 2011;121(5):1953-1960. doi:10.1172/JCI44032

- [28]. Richardson B. Impact of DNA methylation on immune cell function inλ autoimmunity. Clin Immunol. 2007;123(2):133-143. doi: 10.1016/j.clim.2007.06.008
- [29]. DayehT, etal. DNA methylation profiling of pancreatic islets in T1D donors.λ Diabetologia. 2016;59(4):861-871. doi:10.1007/s00125-015-3874-6
- [30]. Ortis F, et al. Cytokine-induced epigenetic changes in β-cells. Cell Death Dis.λ 2010;1: e25. doi:10.1038 / cddis.2010.6
- [31]. Eizirik DL, et al.  $\beta$ -cell apoptosis in T1D: molecular mechanisms. Diabetes. $\lambda$  2009;58(11):2409-2416. doi:10.2337/db09-1150
- [32]. AuneTM, etal. Histone modifications in autoimmune inflammation. Frontλ Immunol. 2017; 8:909. doi:10.3389 / fimmu.2017.00909
- [33]. WangW, etal. Epigenetic regulation of insulin gene expression in diabetes.λ Endocrinology. 2018;159(2): 812-824. doi: 10.1210 / en.2017-00726
- [34]. Pasquali L, et al. SWI/SNF chromatin remodeling in pancreatic  $\beta$ -cell function. $\lambda$  Nat Genet. 2014;46(10):1039-1047. doi:10.1038/ng.3097
- [35]. Joglekar MV, et al. MicroRNA regulation in T1D inflammation. Diabetes Resλ Clin Pract. 2017; 133:53-63. doi: 10.1016/j.diabres.2017.06.005
- [36]. HewardJA, Lindsay MA. Long non-coding RNAs in autoimmunity. Cell Molλ Immunol. 2014;11(2):126-134. doi:10.1038/cmi.2013.59
- [37]. ZhaoM, etal. ncRNA interactions in diabetes pathogenesis. Front Endocrinolλ (Lausanne). 2018; 9:585. doi:10.3389/fendo.2018.00585
- [38]. KahnSE, etal. Pathophysiology and treatment of type 2 diabetes. J Clin Invest.λ 2014;124(4):1212-1220. doi:10.1172/JCI72269
- [39]. DayehT, etal. Epigenetic regulation of metabolic genes in T2D. Epigenomics.λ 2015;7(5):761-770. doi:10.2217 / epi.15.14
- [40]. LingC, RönnT.Epigenetic changes in skeletal muscle in T2D. Epigenomics.λ 2014;6(6):647-650. doi:10.2217 / epi.14.53

- [41]. VolkovP, et al. DNA methylation changes in pancreatic islets from T2D donors.λ Diabetes. 2017;66(4):1033-1045. doi:10.2337/db16-1323
- [42]. El-Osta A, et al. Epigenetic basis of metabolic memory in diabetes. Nat Revλ Endocrinol. 2012;8(7):418-429. doi:10.1038/nrendo.2012.57
- [43]. SunW, etal. Histone deacetylases and insulin resistance. Front Endocrinolλ (Lausanne). 2015; 6:50. doi:10.3389 / fendo.2015.00050
- [44]. Sathishkumar C, et al. Histone modifications and glucose metabolism. Jλ Endocrinol. 2018;236(2): R123-R134. doi:10.1530/JOE-17-0535
- [45]. KazemiR, et al. Role of histone methyltransferases in insulin resistance. Clinλ Epigenetics. 2017; 9:69. doi:10.1186/s13148-017-0360-4
- [46]. PoyMN, etal. MicroRNAs in pancreatic islet biology and diabetes. Ann N Yλ Acad Sci. 2011; 1243:51-59. doi:10.1111/j.1749-6632.2011. 06385.x
- [47]. YangX, etal. Long non-coding RNAs in metabolic regulation. Trendsλ Endocrinol Metab. 2016;27(7):453-462. doi: 10.1016/j.tem.2016.03.010
- [48]. ZhangY, et al. Interactions of ncRNAs and epigenetics in diabetes. Frontλ Endocrinol (Lausanne). 2020; 11:601279. doi:10.3389/fendo.2020.601279
- [49]. Barker DJ. Developmental origins of adult health and disease. J Epidemiolλ Community Health. 2004;58(2): 114-115. Doi: 10.1136/jech.2003.015094
- [50]. Heijmans BT, et al. Persistent epigenetic differences associated with prenatal exposure. Proc Natl Acad Sci U S A. 2008;105(44):17046-17049. doi:10.1073 / pnas. 0806560105
- [51]. P.P. Deepika et al. Formulation and evaluation of doxorubicin nanopsonges for targeted drug delivery African J. Biol. Sci. (South Africa) (2024)
- [52]. Ramu B. Formulation of Lamotrigine Orodispersible Tablets by Using New Generation Super disintegrants Generation Super disintegrants World Journal of Pharmacy and Pharmaceutical Sciences. 2015; 4:631-43
- [53]. S. R. B. B. Saitulasi Ch, Review of Regulatory Aspects Involved in Marketing Authorization of Medical

- Devices., Journal of Advanced Pharmaceutical Research. , 2016.
- [54]. Barrès R, et al. Exercise induces DNA demethylation in skeletal muscle. Cellλ Metab. 2012;15(3):405-411. doi: 10.1016/j.cmet.2012.01.001
- [55]. Ramu, B., Manasa, M.S. (2015). Formulation and Evaluation of Colon Specific Drug Delivery of Press Coated Lansoprazole Tablets. Indo American Journal of Pharmaceutical Research, 5(4),1522.
- [56]. ManikkamM, etal. Environmental toxicants and epigenetic inheritance of  $\lambda$  diabetes risk. Environ Health Perspect. 2012;120(6): 777-785. doi:10.1289 / ehp. 1104302
- [57]. Skinner MK, et al. Transgenerational epigenetic inheritance and environmentalλ factors. Nat Rev Genet. 2014;15(5):259-270. doi:10.1038/nrg3720
- [58]. ChambersJC, et al. Epigenome-wide association of DNA methylation withλ incident type 2 diabetes. Diabetes. 2015;64(2): 571-581. doi:10.2337/db14-0850
- [59]. Ramu B. Formulation of Lamotrigine Oro dispersible Tablets by Using New Generation Super disintegrants Generation super disintegrants World Journal of Pharmacy and Pharmaceutical Sciences. 2015; 4:631-43.
- [60]. Ramu B, Saibaba SV. Role of community pharmacist in management of anaemia. Pharm Pharmocol Int J. 2018;6(3):216–220. DOI: 10.15406 / ppij. 2018.06. 00178..
- [61]. Somarouthu Venkata Saibaba, Bandameedi Ramu. Role of Community Pharmacist in Management of Anaemia. Open Science Journal of Clinical Medicine. Vol. 6, No. 2, 2018, pp. 5-9.
- [62]. Gopikrishna, A.; Ramu, B.; Srikanth, G.; Rajkamal, B. Formulation of isoniazid sustained release formulation by using Carbopol 934 P. Int. J. Appl. Pharm. Sci. Res. 2016, 1, 60–6Villeneuve LM, et al. Epigenetic targets in diabetes therapy. J Mol Med (Berl).λ 2008;86(4):363-372. doi:10.1007/s00109-007-0274-x
- [63]. MilagroFI, et al. Polyphenols as epigenetic modulators in metabolic diseases.λ Nutr Rev. 2013;71(6):418-433. doi:10.1111/nure.12035

[64]. LiuXS, etal. Editing DNA methylation in the mammalian genome. Cell. $\lambda$  2016;167(1):233-247.e17. doi: 10.1016 / j.cell.2016.08.05